Venoactive Drug Therapy On Treatment Of Venous Ulcer

Venöz Ülser Tedavisinde Venoaktif İlaçlar

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Özet

Venöz ülserler anlamlı morbiditeye sebep olan önemli tıbbi sorunlardır. Primer venöz yetmezliğin ileri evrelerinde veya venöz yetmezliğe bağlı oluşan posttrombotik sendrom sonucunda gelişir. Tedavisi kompresyon, bacak elevasyonu, local yara bakımı, ilaç tedavisi ve altta yatan venöz patolojinin cerrahi veya girişimsel olarak düzeltilmesidir. Diğer tedavi yöntemleri eğer kompresyon tedavisi ile birlikte uygulanırsa başarılı olabilir. Biz burada venöz ülser tedavisinde venoaktif ilaçların etkinliğini bilimsel kanıtlar eşliğinde tartıştık.

Anahtar Kelimeler: Venöz ülser; venoaktif ilaçlar; venöz yetersizli

Abstract

Venous ulcers are important medical problem that caused significant morbidity. They develop as a result of advanced chronic venous insufficiency, caused most frequently by primary venous incompetence or the post thrombotic syndrome. Treatment of venous ulcers include compression therapy, leg elevation, local ulcer and skin care, drug treatment, and correction of the underlying venous pathology with surgical and endovenous interventions. Primary and sine qua non method to cure venous ulcer is compression treatment. Other treatment modalities can be successful if only they have performed with compression treatment. In here we discuss the scientific evidence on efficacy of venoactive drug therapy on treatment of venous ulcer.

Key Words: Venous ulcer; venoactive drug; venous insufficiency

INTRODUCTION

Venous ulcers are the most common leg ulcers, and are important medical problem that caused significant morbidity. They develop as a result of advanced chronic venous insufficiency, caused most frequently by primary venous incompetence or the post thrombotic syndrome. Up to 0.5% of the adult Western population has active ulcers, and 0.6% to 1.4% has healed venous ulcers. The overall incidence of venous ulcers in patients older than 45 years of age is estimated at 3.5 per thousand per year; the incidence of venous ulcers has remained unchanged over the last 20 years. (1-3). "Varicose ulcer", "gravitational ulcer", "stasis ulcer" and "hypostatic ulcer" have been used as synonyms for venous ulcer. It is not recommended that these terms be used as the commonly agreed upon term is "venous ulcer" (4). Some risk factors described for CVI development and consequently venous ulcer include age, sex, family history, race, body weight, occupation and number of pregnancies. The influence of each risk factor is controversial. The prevalence of venous ulcer increases progressively with age regarding gender, most studies show a predominance of women affected. The female: male ratio is variable, ranging from 1.5 : 1 to 10:1.(4)

Primary venous valvular insufficiency or venous hypertension caused by venous obstruction is the essential pathology for venous ulcer development. Underlying mechanics can be summarized as : Venous hypertension has significant consequences on the dermal microcirculation and has a negative impact on capillary system. Erythrocytes and macromolecules extravasate from the dermal capillary into the interstitium, where the red cells are broken down and the hemosiderin is engulfed by macrophages. The breakdown products of red blood cells are chemoattractive to leukocytes by activating the endothelial intracellular adhesion molecules. These leukocytes are responsible from releasing of proteolytic enzymes and free oxygen radicals which trigger inflammatory response then Cytokine and proteinase activation causes endothelium and tissue damage. (2, 4-9)

Treatment of venous ulcers include compression therapy, leg elevation, local ulcer and skin care, drug treatment, and correction of the underlying venous pathology with surgical and endovenous interventions. Primary and sine qua non method to cure venous ulcer is compression treatment. Other treatment modalities can be successful if only they have performed with compression treatment. Previously in venous ulcer therapy venoactive medications such as pentoxifylline, prostaglandin, acetylsalicylic acid and sulodexide had been used. (10-17). In this document, we discuss the scientific evidence on efficacy of venoactive drug therapy on treatment of venous ulcer.

Venoactive drugs are safe and reliable drugs and have been used for a long time in all stages of the chronic venous disease. The venoactive drugs can be defined as a group of naturally occurring or synthetic drugs that act on capillary permeability or venous tone (18). They are listed Table 1 (19). Venoactive drugs have been shown to increase venous tone, increase capillary resistance and reduce capillary filtration, improve lymphatic flow and protection against inflammation (3, 18-20). Thus they relieve subjective symptoms such as pain and cramp

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with decreasing limb odema. However despite widely use of these medications there is not a consensus about their efficiency. In Siena consensus paper; a grade A was assigned to calcium dobesilat, MPFF, and O-(betahydroxyethyl) rutosides (HR)-oxerutins, a grade B to escin and ruscus extracts and a grade C to the remaining VADS (21). Recently published Society for Vascular Surgery and the American Venous Forum guideline suggested that diosmin, hesperidin, rutoside, micronized purified flavonoid fraction or horse chestnut seed extract (escin) must be used in addition to compression therapy for chronic venous disease (Grade 2, Evidence level B) (22). Hesperidin therapy accelerates venous ulcer healing and suggested that Diosmin-Hesperidin might be a useful adjunct to conventional therapy in large and long standing ulcers. In a guideline published in 2006 also reported that oral treatment with micronized purified flavonoid fraction (MPFF) may be a useful adjunct to conventional compression therapy in the treatment of leg ulcers-LEVEL I. (29)

Meanwhile, in a non-randomized study with 25 patients in 2003 with calcium dobesilate, subjective complaints, leg diameters, and ulcer sizes of the patients decreased (30). A study with horse chestnut seed extract used for 12 weeks in 2006 reported that no significant affect on venous ulcer healing (31).

Group	Substance	Origin	
Alpha-benzopyrones	Coumarin	Melilot (Melilotus officinalis)	
		Woodruff (Asperula odorata)	
Gamma-	Diosmin	Citruss spp. (Sophora japonica)	
benzopyrones (flavonoids)	Micronized purified flavonoid fraction	Rutaceae aurantiae	
(llavoliolds)	Rutin and rutosides	Sophora japonica	
	O-(β-hydroxyethyl)-rutosides (troxerutin, HR)	Eucalyptus spp. Fagopyrum spp.	
Saponins	Escin	Horse chestnut seed extracts	
	Ruscus extract	Butcher's broom (Ruscus aculeatus)	
Other plant extracts	Anthocyans	Bilberry (Vaccinium myrtillus)	
	Proanthocyanidins (oligomers)	Red wine leaves extracts, Maritime pine	
	Extracts of Ginkgo, heptaminol and troxerutin	Ginkgo biloba	
	Total triterpene fraction	Centella asiatica	
Synthetic products	Calcium dobesilate	Synthetic	
	Benzaron	Synthetic	
	Naftazon	Synthetic	

 Table 1. Classification of the main venoactive drugs (19)
 (19)

Nonetheless effectiveness of these drugs in treatment of venous ulcer still remains controversy. The first study about venoactive drugs in treatment of venous ulcer was reported in 1991 (23). 138 patients with healed venous ulcer were randomized to okserutin and placebo combined with compression sock. Okserutin had no superiority to placebo in terms of venous ulcer recurrence at the end of 3 months therapy. However a guideline which is published in 1995 reported that drug therapy is not beneficial in venous ulcer treatment (24), in 1997 Guilhou et al. reported that 2 months long Diosmin-Hesperidin therapy combined with compression sock decreased ulcer size significantly compared to placebo group in 107 patients who had venous ulcer (25). In a study that is published in 2001, 140 patients treated for 24 weeks by Diosmin-Hesperidin was of significantly well venous ulcer healing (26). Another subsequent study showed that Diosmin-Hesperidin therapy for 6 months accelerates venous ulcer healing (27). Coleridge-Smith et al. reported a meta-analyze including two more study in addition to studies mentioned above in 2005 (28). They have concluded in this meta-analyze that DiosminIn conclusion, Diosmin-Hesperidin combination is the only venotrop that effectiveness is shown in large randomized studies and meta-analyses. Therefore in light of these scientific investigations, American Venous Forum Guideline published in 2009 recommends only Diosmin-Hesperidin among venotrop agents in treatment of venous ulcer as Level of Evidence: A (32). However associate guideline of Society for Vascular Surgery and American Venous Forum published in 2011 reported that micronized purified flavonoid fraction in venous ulcer treatment as Grade 2 – Level of Evidence: B (22), large randomized studies with the other agents could change this knowledge.

REFERENCES

- 1. Gloviczki P, Gloviczki ML. Evidence on efficacy of treatments of venous ulcers and on prevention of ulcer recurrence. Perspect Vasc Surg Endovasc Ther 2009; 21: 259-68
- 2. Etufugh CN, Phillips TJ. Venous ulcers. Clin Dermatol 2007; 25: 121-30
- 3. Nicolaides AN, Allegra C, Bergan J. Management

of chronic venous disorders of the lower limbs: guidelines according to scientific evidence. Int Angiol 2008; 27: 1-59

4. Abbade LP, Lastoria S. Venous ulcer: epidemiology, physiopathology, diagnosis and treatment. Int J Dermatol 2005; 44: 449-56

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- 5. Raffetto JD, Marston WA. Venous ulcer: what is new? Plast Reconstr Surg 2010; 127 (Suppl 1): 279S-288S
- 6. Olivencia JA. Pathophysiology of venous ulcers: surgical implication, review and update. Dermatol Surg 1999; 25: 880-5
- Coleridge-Smith P, Thomas P, Scurr JH, Dormandy JA. Causes of venous ulceration: a new hypothesis. Br Med J 1988; 296: 1726–1727
- Claudy AL, Mirshahi M, Soria C, Soria J. Detection of undegraded fibrin and tumor necrosis factor alpha in venous leg ulcers. J Am Acad Derm 1991; 25: 623– 627
- 9. Powell CC, Rohrer MJ, Barnard MR, Peyton BD, Furman MI, Michelson AD. Chronic venous insufficiency is associated with increased platelet and monocyte activation and aggregation. J Vasc Surg 1999; 30: 844–851.
- De Sanctis MT, Belcaro G, Cesarano MR, Ippolito E, Nicolaides AN, Incandela L, et al. Treatment of venous ulcers with pentoxifylline: a 12-month, doubleblind, placebo controlled trial. Microcirculation and healing. Angiology 2002; 53 (Suppl 1): S49-S51
- 11. Falanga V, Fujitani RM, Diaz C, Hunter G, Jorizzo J, Lawrence PF, et al. Systemic treatment of venous leg ulcers with high doses of pentoxifylline: efficacy in a randomized, placebo-controlled trial. Wound Repair Regen. 1999; 7: 208-13
- 12. Dale JJ, Ruckley CV, Harper DR, Gibson B, Nelson EA, Prescott RJ. Randomized, double blind placebo controlled trial of pentoxifylline in the treatment of venous leg ulcers. BMJ. 1999 2; 319: 875-8
- 13. Ferrara F, Meli F, Raimondi F, Amato C, Bonura F, Mulè G, et al. The treatment of venous leg ulcers: a new therapeutic use of iloprost. Ann Surg 2007; 246: 860-5
- 14. Layton AM, Ibbotson SH, Davies JA, Goodfield MJ. Randomized trial of oral aspirin for chrinic venous leg ulcers. . Lancet 1994; 344: 164-5
- 15. Milio G, Mina C, Cospite V, Almasio PL, Novo S. Efficacy of the treatment with prostaglandin E-1 in venous ulcers of the lower limbs. J Vasc Surg 2005;42:304-8.
- Werner-Schlenzka H, Kuhlmann RK. Treatment of venous leg ulcersvwith topical iloprost: a placebo controlled study. VASA 1994; 23:145- 50
- 17. Coccheri S, Scondotto G, Agnelli G, Aloisi D, Palazzini E, Zamboni V. Randomised, double blind, multicentre, placebo controlled study of sulodexide in the treatment of venous leg ulcers. Thromb Haemost 2002;87:947-52.
- 18. Gohel MS, Davies AH. Pharmacological agents in the treatment of venous disease: An update of the available evidence. Curr Vasc Pharmacol 2009; 7:

- 303-8
 19. Perrin M, Ramelet AA. Pharmacological treatment of primary chronic venous disease: Rationale, results and unanswered questions. Eur J Vasc Endovasc Surg 2011; 41: 117-25
- 20. Boisseau MR. Pharmacological targets of grugs employed in chronic venous and lymphatic insufficiency. Int Angiol 2002; 21: 33-9
- 21. Ramelet AA, Boisseau MR, Allegra C, et al. Venoactive drugs in the management of chronic venous disease. An international consensus statement: current medical position, nprospective views and final resolution. Clin Hemorheol Microcirc 2005; 33: 309-19
- 22. Gloviczki P, Comerota AJ, Dalsing MC, et al. The care of patients with varicose veins and associated chronic venous diseases: clinical practice guidelines of the Society for Vascular Surgery and the American Venous Forum. J Vasc Surg. 2011; 53 (5 Suppl): 2S-48S.
- 23. Wright DD, Franks PJ, Blair SD, Backhouse CM, Moffatt C, McCollum CN. Oxerutins in the prevention of recurrence in chronic venous ulceration: randomized controlled trial. Br J Surg 1991; 78: 1269-70
- 24. Douglas WS, Simpson NB. Guidelines fort he management of chronic venous leg ulceration. Report of a multidisciplinary workshop. Br J Dermatol 1995; 132: 446-52
- 25. Guilhou JJ, Dereure o, Marzin L, et al. Efficacy of Daflon 500 mg in venous leg ulcer healing: a doubleblind, randomized, controlled versus placebo trial in 107 patients. Angiology 1997; 48: 77-85
- 26. Glinski W, Chodynicka B, Roszkiewicz J, et al. Effectiveness of a micronized purified flavonoid fraction (MPFF) in the healing process of lower limb ulcers. An open multicentre study, controlled and randomized. Minerva Cardioangiol 2001; 49: 107-14
- 27. Roztocil K, Stvrtinova V, Strejcek J. Efficacy of a 6-month treatment with Daflon 500 mg in patients with venous leg ulcers associated with chronic venous insufficiency. Int Angiol 2003; 22: 24-31
- 28. Coleridge-Smith P, Lok C, Ramelet AA. Venous leg ulcer: a meta-analysis of adjunctive therapy with micronized purified flavonoid fraction. Eur J Vasc Endovasc Surg 2005; 30: 198-208
- 29. Robson MC, Cooper DM, Alsam R. Guidelines for the treatment of venous ulcers. Wound Rep Reg 2006; 14: 649-62
- 30. Kaur C, Sarkar R, Kanwar AJ, Attri AK, Dabra AK, Kochhar S. An open trial of calcium dobesilate in patients with venous ulcers and stasis dermatitis. Int J Dermatol 2003; 42: 147-52
- 31. Leach MJ, Pincombe J, Foster G. Clinical efficacy of horsechestnut seed extract in the treatment of venous ulceration. J Wound Care 2006; 15: 159-67
- 32. Moneta GL, Partsch H. Compression therapy for venous ulceration. In: Gloviczki P, ed. Handbook of Venous Disorders . 3rd ed. London, UK: Hodder Arnold; 2009:348-358

In vitro Investigation of the Effect of Cinnamon on Protein Glycosylation, Na+-K+ ATPase, Ca++ ATPase and Lipid Peroxidation in Human Erythrocytes Exposed to High Glucose Concentration

Tarçının (Cinnamon) Yüksek Glukoz Konsantrasyonlarına Maruz Bırakılan İnsan Eritrositlerinde (in vitro) Protein Glikozilasyonu, Na+-K+ ATPaz, Ca++ ATPaz ve Lipid Peroksidasyonu Düzeylerine Etkisinin Araştırılması

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Özet

Abstract

Amaç: Bu çalışmada, yüksek glukoz konsantrasyonuna maruz bırakılan insan eritrositlerinde tarçının protein glikozilasyonu, Na+-K+ ATPaz ve Ca++ ATPaz aktiviteleri ve lipid peroksidasyonu seviyelerine etkisi in vitro olarak araştırılmıştır.

Gereç ve Yöntem: Bu amaçla, sağlıklı bireylerden alınan kan örneklerini ya normal glikoz (kontrol grubu) ya da yüksek glikoz konsantrasyonlarına maruz bırakılmış ve daha sonra farklı konsantrasyonlarda tarçın ile inkübe edilmiştir.

Bulgular: Yüksek glukoz konsantrasyonuna maruz bırakılan eritrosit örneklerinde Na+-K+ ATPaz ve Ca++ ATPaz aktiviteleri kontrol grubuna göre daha düşük bulundu ve bu iki grup arasındaki fark istatistiksel olarak anlamlıydı (p <0.001). Tarçına maruz bırakılan gruplarda tarçın konsantrasyonlarının artması nedeniyle bu iki zar enziminin aktiviteleri istatistiksel olarak önemli bir seviyede artmıştır. Ayrıca MDA ve HbA1c düzeylerinin yüksek glikoz grubunda normal glikoz grubuna göre daha fazla arttığı ve tarçının etkisi altında, tarçın konsantrasyonuna bağlı olarak azaldığı tespit edildi. Sonuç olarak, tarçın Na+-K+ ATPaz ve Ca++ ATPaz aktivitelerini arttırdı ve yüksek glikoz konsantrasyonuna maruz bırakılan eritrositlerdeki lipid peroksidasyonu seviyesini düşürdü, normal glikoz konsantrasyonlarında bunu yaptı.

Sonuç: Tarçının bu parametreler üzerindeki etkilerinin, yüksek kan şekeri seviyesi ile karakterize edilen ve dünyada yaygın olarak bilinen bir hastalık olan diabetes mellitus üzerinde özel bir öneme sahip olduğu sonucuna varılmıştır.

Anahtar Kelimeler: Anahtar kelimeler: Tarçın, Na+-K+ ATPaz, Ca++ ATPaz, Lipid Peroksidasyonu, Eritrosit Membranı, Protein Glikozilasyonu. **Objective:** In this study, the effect of cinnamon on protein glycosylation, Na+-K+ ATPase and Ca++ ATPase activities and lipid peroxidation levels in human erythrocytes exposed to high glucose concentration was investigated in vitro.

Materials and methods: For this purpose, the blood samples obtained from healthy individuals were exposed to either normal glucose (control group) or high glucose concentrations and then incubated with cinnamon at different concentrations.

Results: In samples of erythrocyte, which have been exposed to high glucose concentration, Na+-K+ ATPase and Ca++ ATPase activities were found to be lower than those of control group and the differences between these two groups were statistically significant (p<0.001). In the groups exposed to cinnamon, the activities of these two membrane enzymes increased at a statistically significant level due to the increase of cinnamon concentrations. It was also detected that MDA and HbA1c levels increased more in high glucose group than normal glucose group, and they also decreased under the effect of cinnamon, directly depending on the concentration of applied cinnamon. As a result, cinnamon increased the activities of Na+-K+ ATPase and Ca++ ATPase and decreased the level of lipid peroxidation in erythrocytes subjected to high glucose concentration, did so in normal glucose concentrations.

Conclusion: It is concluded that the effects of cinnamon on these parameters have a special importance on diabetes mellitus, a disease known commonly all over the world, which is characterized by high blood glucose level.

Key Words: Key Words: Cinnamon, Na+-K+ ATPase, Ca++ ATPase, Lipid Peroxidation, Erythrocyte Membrane, Protein Glycosylation.

INTRODUCTION

There are many plants in the nature that are considerably valuable for the fields of medicine and chemistry. In recent years, taking advantage of these plants for the purpose of protection from diseases and finding cures have become more essential. This situation implies the necessity for conscientious studies on such plants. Diabetes Mellitus (DM) is an endocrinal and metabolic disease that arises from absolute or relative insulin deficiency or insulin resistance, and is characterized by alterations in carbohydrate, lipid and protein metabolisms (1). During the course of this disorder, specific complications such as retinopathy, nephropathy, and atherosclerosis may develop, and each year, thousands of people around the world die due to these diabetic complications (2). It has been reported

This study was published as a Master Degree Thesis in KSU in 2006 and as a poster in 2nd International Congress on Cell Membranes and Oxidative Stress: Focus on Calcium Signalling and TRP Channels.

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that in patients with diabetes, there is an increase in the production of free oxygen radicals and lipid peroxidation by-products, and the antioxidant defense system becomes insufficient (3). Increases in the production of free radicals contribute to the emergence and advancement of diabetic complications (4). In diabetes, reactive oxidants that cause oxidative damage are created due to the high level of glucose (5). Non-enzymatic glycosylation, metabolic stress caused by the changes in the energy metabolism, sorbitol tract activity, levels of inflammatory mediators and localized tissue damage caused by changes in the antioxidant defense system are the mechanisms that increase oxidative stress in diabetic patients (4).

The role of Na⁺ ions in the transportation of glucose within the cells is well documented and glucose uptake is mediated by glucose-Na⁺ symport system (6). In connection with the latter, Na^+-K^+ ATPase is an enzyme that is responsible for the efflux and influx of Na⁺ and K^+ ions, respectively (6). In addition, Ca^{+2} ATPase, which is responsible for the efflux of Ca⁺² ions, together with Na⁺-K⁺ ATPase plays a central role in keeping the intracellular and extracellular concentrations of these ions within the physiological levels (7). Hyperglycaemia and consumption of glucose within the cell cause changes in intracellular and extracellular concentrations of Na⁺ ions directly and Ca++ ions indirectly (7), and hence it may be implied that the Na⁺-K⁺ ATPase and Ca⁺⁺ ATPase enzyme activities could be affected by these changes. In previous studies, it has been indicated that impaired sodium transport and Na⁺-K⁺ ATPase activity in the erythrocyte membrane possibly have prominent roles in the pathophysiology of chronic complications of diabetes mellitus (8-11). Furthermore, it has been reported in several studies conducted in vivo and in vitro conditions that there are changes in the Na⁺-K⁺ ATPase and Ca⁺⁺ ATPase activities in accordance with the increase in glycaemia (9, 10). Moreover, it has been reported that various plants, including Cinnamon, may improve the glucose metabolisms in diabetic patients due to their hypoglycemic effects (12).-

Cinnamon is a fragrant evergreen tree belonging in the lauraceae family which is native to the South and South East Asia (13). Its leaves are leathery and have usually three veins (13). It has a strong and special scent, and a sweet and hot taste (13). Its bark has been used as spice and medicine for thousands of years (13). Especially in the Far East and Indian cultures, it has been used for the treatment of indigestion, loss of appetite, bloating, and stomach aches (14). After so many years of traditional usage, there have been various scientific studies on animals in order to utilize this plant in modern therapy methods (14). Cinnamon is obtained from the bark of Cinnamomi cassia tree (15). It contains cinnamic aldehyde, cinnamic acid, tannin and methyl hydroxyl chalcone polymers (MHCP) (15). There are two main species found in nature: Cinnamomum Zeylanicum N. and Cinnamomum Verum (16). Additionally, there is 1-4% aromatic volatile oil in the composition of cinnamon (17). This oil contains ethyl cinnamate, eugenol, cinnamaldehyde, beta-caryophyllene, linalool, and methyl chavicol (17). It is a plant which is frequently mentioned in studies on diabetes with its hypoglycemic effects; it has been reported that the cinnamon extracts

increase the in vitro ingestion of glucose and glycogen synthesis, and it has positive effect on the glucose metabolism by increasing the phosphorylation of insulin receptors (12). Moreover, it is also reported that cinnamon has antioxidant effects (18). It has been shown that cinnamon decreases the glucose level in blood and this effect is related to the methyl hydroxyl chalcone polymer (MHCP) that is found in cinnamon. It has been indicated that this substance shows effects similar to that of insulin. MHCP is an important biological compound which has anti-bacterial, anti-fungal and fever reducing properties, as well as having the effect of reducing the cholesterol in the blood (12, 19). Especially in United States and Germany, cinnamon is used as the active ingredient in anti-flatulent herbal drugs. It also has a role in the treatment of cold and inflammatory diseases (20). A significant benefit of phytochemicals, vitamins, minerals, and fruits and vegetables in foods is thought to be capable of scavenging free radicals, lowering the incidence of chronic diseases (2).

The aims of this study was determined the effect of cinnamon on protein glycosylation, Na^+-K^+ ATPase and Ca^{++} ATPase activities and lipid peroxidation levels in human erythrocytes exposed to high glucose concentration.

MATERIAL AND METHODS

Sampling and Study groups

Erythrocytes used in the study were obtained from blood samples collected from healthy and voluntary subjects. During the study, a portion of the samples was exposed to 45 mM glucose and the rest was exposed to 6 mM glucose. Afterwards, these samples were incubated in different concentrations of cinnamon. Glucose applied to the erythrocyte samples were grouped according to the cinnamon concentrations as stated below. Study groups are shown in Table I.

Incubation of Erythrocytes in Glucose and Cinnamon

Heparinized blood samples were isolated from plasma by means of centrifugation (4500 x g). Erythrocytes were brought to 10% hematocrit with the saline solution buffered with phosphate (6). Then, same amounts (100 µL) of these erythrocytes were added glucose solutions and slowly stirred so that the final concentration is 6mM for the normal concentration glucose group and 45 mM for the high concentration group. 10 µL of antibiotics (sefazol) were added to the incubation medium in order to prevent the reproduction of microorganisms (7). Following this procedure, different concentrations of cinnamon were added to the samples, and incubated for 1 hour, 24 hours and 48 hours at 37° C temperature. The 1 hour incubated samples were used in order to determine the enzyme activity, and 1, 24 and 48 hour incubated samples were used for the MDA and HbA₁, measurements.

Separating the Erythrocyte Membrane

The erythrocyte membrane was separated by

Table I. Study groups of the present study

Erythrocytes exposed to 45 mM glucose (Experimental groups)	Erythrocytes exposed to 6 mM glucose (Control groups)
Group IA: High concentration glucose medium (45 mM glucose)	Group IB: Normal concentration glucose medium (6 mM glucose)
Group IIA: 45 mM glucose + 50 μM Cinnamon	Group IIB: 6 mM glucose + 50 μM Cinnamon
Group IIIA: 45 mM glucose + 100 µM Cinnamon	Group IIIB: 6 mM glucose + 100 μM Cinnamon
Group IVA: 45 mM glucose + 100 μM Cinnamon (24 h incubation)	Group IVB: 6 mM glucose + 100 μ M Cinnamon (24 h incubation)
Group VA: 45 mM glucose + 100 μM Cinnamon (48 h incubation)	Group VB: 6 mM glucose + 100 µM Cinnamon (48 h incubation)
Group VIA: 45 mM glucose + 200 µM Cinnamon	Group VIB: 6 mM glucose + 200 µM Cinnamon

using Moretti method (21). The obtained erythrocyte membranes were used within the same day in order to determine the Na⁺-K⁺ ATPase and Ca⁺⁺ ATPase activities.

Measurement of Na⁺-K⁺ ATPase Enzyme Activity

 Na^+-K^+ ATPase enzyme activity was measured based on the Mazzanti method. 1 mL incubation medium was used in order to determine the enzyme activity. The experimental procedure was then repeated without the ouabain in the medium. The activity was calculated by subtracting the Pi value measured with the oubaine from the Pi value measured without the oubaine. The enzyme activity is given as the amount of inorganic phosphate produced from ATP by 1 mg protein in 1 hour (µmol Pi/ mg protein.hour) (22).

Measurement of Ca⁺⁺ ATPase Enzyme Activity

Ca⁺⁺ ATPase enzyme activity was measured based on the Flecha method. 1 mL incubation medium was used in order to determine the enzyme activity. The enzyme activity is given as the amount of inorganic phosphate produced from ATP by 1 mg protein in 1 hour (μ mol Pi/ mg protein.hour) (23).

Determination of the Amount of Inorganic Phosphate

The amount of inorganic phosphate was measured by modifying the Ames method. This method was essentially based on the principle of reduction of molybdate complex by ascorbic acid (24).

Determination of Lipid Peroxidation

Determination of lipid peroxidation is based on the principle of spectrophotometric evaluation of the absorbance of the pink-red color resulting from reaction between the products (mainly MDA) and TBA (25, 26). The net absorbance is calculated by subtracting the baseline absorbance from the absorbance of the sample. The MDA concentration is calculated in terms of nmol/ mL by making use of the molar extinction coefficient of MDA-TBA complex in 532 nm (1.56x10⁵ M⁻¹ cm⁻¹) and taking dilution factor into consideration.

Determination of Total Protein

Copper-protein complex is created in alkali solution. This complex reduces the phosphomolybdatephosphotunstate reactive and it gives a dark blue color. The intensity of this dark blue is directly proportional to the concentration of protein in the medium. One important aspect to watch out during the addition of folinic reactive is that this reactive is durable only in acidic medium. The described reduction process takes place in pH 10. Therefore the folinic reactive should be immediately added to the copper-protein solution and be immediately stirred strongly. Thus, the reduction process can take place before the decomposition of phosphomolybdatephosphotunstate reactive (27). Albumin is used as the standard solution.

Statistical Analysis

All values are expressed as mean \pm standard deviation (standard error of mean (S.E.M.) is better!). One way analysis of variance (ANOVA) was carried out in order to determine whether there had been significant difference between the groups, and TUKEY HSD test, which is a POST HOC test, was utilized in order to compare the groups in pairs between themselves. Limit of significance is taken as p<0.05. These statistical procedures were carried out by the use of SPSS package program (version 13.0).

RESULTS

In this study, the effect of cinnamon on the levels of glycosylated hemoglobin (HbA_{1c}) and MDA, and activities of erythrocyte membrane Na^+-K^+ ATPase and Ca⁺⁺ ATPase in human erythrocytes which have been exposed to normal and high concentration of glucose; and the obtained results are statistically analyzed.

Erythrocyte Membrane Na⁺-K⁺ ATPase and Ca⁺⁺ ATPase Activities, MDA and HbA_{1c} Values

Erythrocyte membrane ATPases activities, MDA and HbA_{1c} values in human erythrocytes which have been exposed to normal and high concentration of glucose and then applied different concentrations of cinnamon are shown in Table II.

As shown in Table II, in this study, in the human

erythrocytes that are *in vitro* exposed to 1 hour of high concentration of glucose, the ATPases activities are

Erythrocyte Membrane Na⁺-K⁺ ATPase and Ca⁺⁺ ATPase Activities, MDA and HbA_{1c} Values under

Table II. Erythrocyte membrane ATPases, erythrocyte membrane MDA and HbA_{1c} values in mediums with normal and high glucose concentration (X±SD; Mean±Standard Deviation)

CINNAMON	Na ⁺ K ⁺ ATPase (μmol Pi/mg protein. hour)	Ca ⁺⁺ ATPase (µmol Pi/mg protein. hour)	MDA (nmol/ gr hemoglobin)	HbA _{1c} (%)
45 mM Glucose $(n = 5)$	0.0408±0.0003	0.0679±0.0003	1.1836±0.0211	7.4200±0.1303
$45 \text{ mM Glucose} + 50 \mu \text{M cinnamon} (n = 5)$	0.0422±0.0002	0.0734±0.0002	0.9836±0.0198	7.0400±0.1140
$45 \text{ mM Glucose} + 100 \mu\text{M cinnamon} (n = 5)$	0.0439±0.0003	0.0748±0.0002	0.9096±0.0176	6.7400±0.1516
$\begin{array}{c} 45 \text{ mM Glucose} + 200 \ \mu\text{M cinnamon} (n = 5) \end{array}$	0.0470±0.0002	0.0784±0.0001	0.7666±0.0237	6.4400±0.2073
6 mM Glucose (n =5)	0.0524±0.0003	0.0788±0.0003	0.8830±0.0209	5.7000±0.1581
6 mM Glucose + 100 μ M cinnamon (n = 5)	0.0575±0.0002	0.0809±0.0005	0.8068±0.0167	5.5600±0.1816

Table III. Erythrocyte membrane ATPase activities of groups on which cinnamon is applied ($X \pm SD$; Mean \pm Standard Deviation)

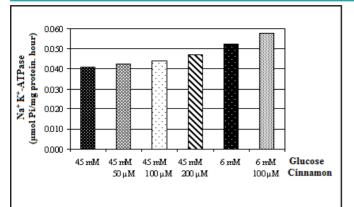
Groups	Na ⁺ -K ⁺ ATPase (μmol Pi/mg protein. hour)	Ca ⁺⁺ ATPase (µmol Pi/mg protein. hour)
Group I (45 mM Glucose)	0.0408±0.0003	0.0679±0.0003
Group II (45 mM Glucose + 50 μM cinnamon)	0.0422±0.0002	0.0734±0.0002
Group III (45 mM Glucose + 100 μM cinnamon)	0.0439±0.0003	0.0748±0.0002
Group IV (45 mM Glucose + 200 μM cinnamon)	0.0470±0.0002	$0.0784 {\pm} 0.0001$
Group V (6 mM Glucose)	0.0524±0.0003	0.0788±0.0003
Group VI (6 mM Glucose + 100 μM cinnamon)	0.0575±0.0002	0.0809±0.0005

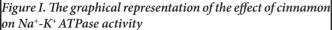
lower than that of the control group (Group V) which is exposed to normal concentration of glucose; and the MDA and HbA_{1c} values are found to be higher than the control group. In groups where cinnamon has been applied, it has been detected that there is an increase in the activity of these enzymes and decrease in the levels of MDA and HbA_{1c}, depending on the concentration of the applied cinnamon. In the study, it has been determined from the trials that the most ideal cinnamon incubation duration for the erythrocyte samples in normal and high concentration glucose medium is 1 hour; and hence the erythrocytes have been incubated with cinnamon for 1 hour.

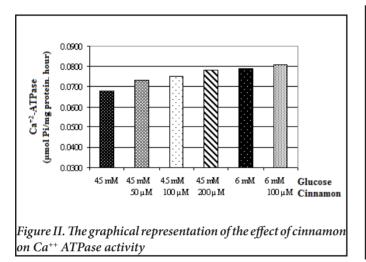
Cinnamon Application

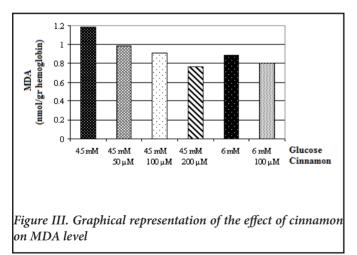
Erythrocyte membrane ATPases activities, MDA and HbA_{1c} values in human erythrocytes which have been exposed to normal and high concentration of glucose and then applied different concentrations of cinnamon are shown in Table IV and Table V, Figure I, Figure II, Figure III and IV and the statistical comparison of the obtained values are shown in Table IV.

In this study, in human erythrocytes, the Na⁺-K⁺ ATPase and Ca⁺⁺ ATPase activity values are found that are exposed *in vitro* to high glucose for 1 hour, to be lower than those of the control group (Group V) with a

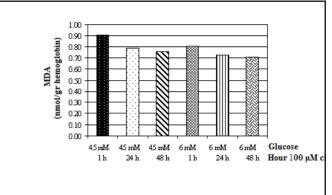


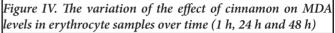


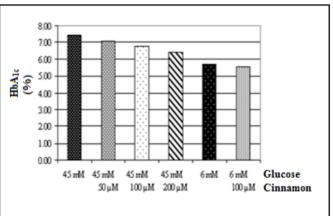


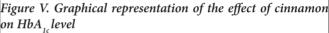


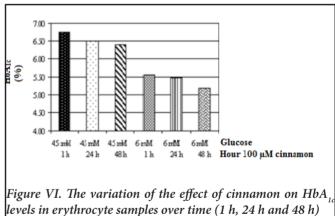
statistical significance (p<0.001) (Table III). In groups which cinnamon has been applied, it has been observed that the levels of activities of these enzymes are higher with statistical significance. In the group that is exposed to 45mM glucose, the Ca⁺⁺ ATPase activity levels are observed to rise to the levels of the control group under the application of 200 μ M cinnamon. In addition, a statistically significant (p<0.001) increase has been detected in both enzyme activities in the 6 mM glucose medium under the effect of 100 μ M cinnamon. The study indicates that in high and normal glucose concentrations, the cinnamon has the effect of increasing the erythrocyte membrane ATPases activities. It has been noted that there is statistically significant (p<0.001) variance among











groups except for groups IV-V for $Na^{\scriptscriptstyle +}\text{-}K^{\scriptscriptstyle +}$ ATPase and $Ca^{\scriptscriptstyle ++}$ ATPase.

In this work, it has been detected that there is a statistically significant increase in MDA and HbA_{1c} values due to the effect of the high concentration glucose compared to the control group (Group V) on which the normal concentration of glucose is applied; and with the effect of cinnamon, MDA and HbA_{1c} values have been lowered depending on the concentration of the applied cinnamon (Table IV, V). In order to determine the effect of cinnamon on the lipid peroxidation and glycosylation in the erythrocytes in normal and high concentration glucose mediums, MDA and HbA_{1c} levels have been measured during the 24 and 48 hours following the cinnamon incubation. Comparison of MDA and HbA_{1c}

Guleșci Table IV. Erythrocyte membrane MDA v	values in groups on which cinna	tmon is applied ($X \pm SD$; Mea	an ± Standard Deviation)
Groups	MDA (nmol/gr hemoglobin)	Compared Groups	*p
Group I (45 mM Glucose)	1.1836±0.0211	p<0.001	
Group II (45 mM Glucose + 50 μM cinna- mon)	0.9836±0.0198	p<0.001	
Group III (45 mM Glucose + 100 μM cinna- mon)	0.9096±0.0176	III-V	0.526
Group III-A (45 mM Glucose + 100 μM cinna- mon) (24 h incubation)	0.7860±0.0114	III.A-III.B III.A-IV III.A-VI III.A-VI.A	0.578 0.865 0.812 0.002
Group III-B (45 mM Glucose + 100 μM cinna- mon) (48 h incubation)	0.7604±0.0312	III.B-IV III.B-VI III.B-VI.A III.B-VI.B	1.000 0.020 0.298 0.003
Group IV (45 mM Glucose + 200 μM cinna- mon)	0.7666±0.0237	IV-VI IV-VI.A IV-VI.B	0.071 0.113 0.001
Group V (6 mM Glucose)	0.8830±0.0209	p<0.001	
Group VI (6 mM Glucose +100 μM cinna- mon)	0.8068±0.0167	p<0.001	
Group VI-A (6 mM Glucose + 100 μM cinna- mon) (24 h incubation)	0.7290±0.0110	VI.A-VI.B	0.651
Group VI-B (6 mM Glucose + 100 μM cinna- mon) (48 h incubation)	0.7048±0.01658	p<0.001	

values obtained from 1 hour (group III), 24 hours (group III-A) and 48 hours (group III-B) incubations of 100 μ M cinnamon under high glucose concentration, indicate that lipid peroxidation and hemoglobin glycosylation have been reduced with the duration of application of cinnamon. Similarly, comparison of MDA and HbA_{1c} values obtained from 1 hour (group III), 24 hours (group III-A) and 48 hours (group III-B) incubations of 100 μ M cinnamon under normal glucose concentration, indicate that lipid peroxidation and hemoglobin glycosylation have been reduced with the duration of application for μ M cinnamon under normal glucose concentration, indicate that lipid peroxidation and hemoglobin glycosylation have been reduced with the duration of application

of cinnamon. In the following tables, each group was compared with the other groups and their subgroups.

It has been detected that in terms of MDA values, the variance among groups on which cinnamon is applied are statistically significant (p<0.001) except for group III-V, group III.A-III.B, group III.A-IV, group III.A-VI, group III.B-IV, group III.B-VI.A, group IV-VI, group IV-VI.A and group VI.A-VI.B. Moreover, in terms of HbA_{1c} values, it is also statistically significant (p<0.05) except for group II-III, group III-III.A, group III-III.B, group III-IV, group III.A-III.B, group III-IV, group Güleşci

Effect of cinnamon

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Groups	HbA _{1c} (%)	Compared Groups	*p
Group I (45 mM Glucose)	7.4200±0.1303	I-II	0.021
Group II (45 mM Glucose + 50 μM cinnamon)	7.0400±0.1140	II-III	0.137
2		III-III.A	0.395
Group III (45 mM Glucose + 100 μM cinnamon)	6.7400±0.1516	III-III.B	0.057
		III-IV	0.137
Group III-A (45 mM Glucose + 100 μM cinnamon)	6.5000±0.1581	III.A-III.B	0.992
(45 mM Glucose + 100 µM cinnamon) (24 h incubation)		III.A-IV	1.000
Group III-B (45 mM Glucose + 100 μM cinnamon) (48 h incubation)	6.4000±0.0707	III.B-IV	1.000
		· · · · · · · · · · · · · · · · · · ·	
Group IV (45 mM Glucose + 200 μM cinnamon)	6.4400±0.2073	p<0.0)5
		V-VI	0.933
Group V (6 mM Glucose)	5.7000 ± 0.1581	V-VI.A	0.517
		V-VI.B	0.001
Group VI	5 5600+0 1816	VI-VI.A	0.999
(6 mM Glucose +100 µM cinnamon)	5.5600±0.1816	VI-VI.B	0.035
Group VI-A (6 mM Glucose + 100 μM cinnamon) (24 h incubation)	5.4800±0.2387	VI.A-VI.B	0.203
Group VI-B (6 mM Glucose + 100 μM cinnamon) (48 h incubation)	5.2000±0.1581	p<0.05	

III.B-IV.B, group V-VI, group V-VI.A, group VI-VI.A and group VI.A-VI.B.

DISCUSSION

Cinnamon is recently a popular plant in diabetes which is known for its hypoglycemic effects, and it has been reported that the cinnamon extracts increase *in vitro* glucose inhibition and glycogen synthesis, and have positive effects on the glucose metabolism by increasing the phosphorilization of insulin receptors (12). It has also been stated that cinnamon has antioxidant effects (18).

Some researchers indicate that cinnamon may have antidiabetic effect by way of reducing the glucose level in the blood (15, 28). This study shows that cinnamon has beneficial effects in high glucose mediums.

Jain et al. have investigated the effects of lipoic acid which beneficial to the human body, on the ATPases activities in glycosylated human erythrocytes. For this purpose, erythrocyte membranes prepared from the blood samples of healthy individuals have been incubated with 0.2 mM lipoic acid under normal (6 mM) and high (45 mM) concentrations of glucose. As a result, it has been observed that the lipoic acid has increased the activity of both enzymes in high glucose membranes, and has no effect on enzyme in normal glucose membranes. In the same work, it has also been stated that under high glucose concentrations, lipoic acid prevents lipid peroxidation and protein glycosylation (6).

Similar finding have also been obtained by

Nandhini et al. in erythrocytes subjected to high concentrations of glucose with taurine (29).

CONCLUSIONS AND SUGGESTIONS

The following conclusions are drawn from this work where effects of cinnamon on the erythrocyte membrane Na⁺-K⁺ ATPase, Ca⁺⁺ ATPase activities and glycosylated hemoglobin (HbA_{1c}) and MDA levels in the human erythrocytes that are subjected to *in vitro* high and normal glucose concentrations:

1. The levels of activities of $Na^+-K^+ATPase$ and $Ca^{++}ATPase$ have been found to be lower in the human erythrocytes that are subjected to *in vitro* high glucose concentrations (45mM) than those of control group which is subjected to normal glucose concentration (6mM) at a statistically significant level (p<0.001).

2. It has been detected that for groups which have been exposed to 50, 100 and 200 μ M of cinnamon for 1 hour, the activities of these enzymes have shown a statistically significant increase depending on the concentration of the cinnamon used.

3. In the group that is exposed to 45 mM glucose, it has been observed that under the effect of 200 μ M cinnamon, ATPases activities have increased to the level of the control group.

4. In the presence of 6 mM of glucose, a statistically significant (p<0.001) increase in the activities of both enzymes have been detected under the effect of 100 μ M cinnamon.

5. The study indicates cinnamon increases erythrocyte membrane ATPases activities under normal and high glucose concentrations.

6. The study shows that the due to the high glucose concentration, MDA and HbA_{1c}values have significantly increased compared to the control group on which normal concentration glucose is applied, and in addition, these values decreased under the effect of cinnamon, depending on the concentration of the applied cinnamon.

7. In order to determine the long term effect of cinnamon on lipid peroxidation and glycosylation in erythrocytes in normal and high glucose mediums, the MDA and HbA_{1c}levels have been measured in 24 and 48 hours following the cinnamon incubation. It has been detected that for high and normal glucose concentrations, as a result of 1 h, 24 h and 48h 100 μ M cinnamon incubations, the MDA and HbA_{1c}values have decreased as the duration of incubation increases.

In conclusion, this study indicates that at the end of a 1 hour in vitro duration, cinnamon increases the Na⁺-K⁺ ATPase and Ca⁺⁺ ATPase activity in human erythrocyte membrane and reduces the MDA and HbA_{1c} levels. It has also been detected that these effects of cinnamon are also valid for erythrocytes that are exposed to high concentration of glucose, and these effects increase depending on the concentration of the applied cinnamon.

In the modern world where the living conditions keep getting harder, diabetes mellitus which needs constant monitoring and treatment affects patients with both acute and chronic complications and these complications cause physical as well as mental and social problems for the patients. Increase in life span due to modern treatment methods causes the frequency and variety of diabetes complications and early diagnosis and treatment becomes more and more essential for a healthy living. In diabetes treatment, natural and herbal remedies draw attention in addition to diet, oral antidiabetics and insulin.

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REFERENCES

- American Diabetes Association. Diagnosis and classification of diabetes. Diabetes Care. 2008; 31: S55-S60.
- 2. Dal S, Sigrist S. The Protective Effect of Antioxidants Consumption on Diabetes and Vascular Complications. Diseases. Review, 2016; 4: 1-51.
- King GL, Banskota NK. Mechanisms of diabetic microvascular complications. In: Kahn, C.R, Weir GC. eds, Joslin's Diabetes Mellitus. (International Ed. Thirteenth Ed. Company). 1994; 634-648.
- 4. Maxwell SRJ, Thomason H, Sandler D, Leguen C, Baxter MA, Thorpe GHG, et al. Poor glycaemic control is associated with reduced serum free radical scavenging (antioxidant) activity in non-insulindependent diabetes mellitus. Ann Clin Biochem. 1997; 34: 638-644.
- Orei NN, Zidek W, Tepel M. Increased intracellular generation of reactive oxygen species in mononuclear leukocytes from patients with diabetes mellitus type 2. Exp Clin Endocrinol Diabetes. 2000; 108: 175-80.
- Jain SK, Lim G. Pyridoxine and pyridoxamine inhibits superoxide radicals and prevents lipid peroxidation, protein glycosylation, and Na⁺, K⁺ -ATPase activity reduction in high glucose-treated human erythrocytes. Free Radical Biology & Medicine. 2001; 12: 595-604.
- Jain SK, Lim G. Lipoicacid decreases lipid peroxidation and protein glycosylation and increases (Na⁺, K⁺) and Ca⁺²ATPase activities in high glucosetreated human erythrocytes. Free Radical Biology & Medicine. 2000; 11: 1122-1128.
- Davis FB, Davis PJ, Nat G, Blas SD, Macgillivra M, Gutman S, et al. The effect of *in vivo* glucose administration on human erythrocyte Ca⁺⁺-ATPase activity and on enzyme responsiveness *in vitro* to thyroid hormone and calmodulin. Diabetes. 1985; 34: 639-646.
- Bilgin R. Glukozun Eritrosit Zarlarında Bulunan Na⁺- K⁺ ATPaz ve Ca⁺² ATPaz Enzim Aktivitelerine Etkisinin İn Vitro ve İn Vivo Koşullarda Araştırılması. Çukurova Üniversitesi Doktora Tezi, Adana. 1995; 100p.
- 10. Kızıltunç A, Akçay F, Polat F, Kuşkay S, Şahin YN. Reduced lecithin: Cholesterol acyltransferase

(LCAT) and Na⁺, K⁺ -ATPase activity in diabetic patients. Clinical Biochemistry. 1997; 2: 177-182.

- Gürbilek M, Dağlar C, Aköz C. The effect of disease duration on of erythrocyte membrane Na⁺, K⁺ -ATPase enzyme activity, lipid peroxidation, and DHEA(S), glucose and lipid levels in the diabets mellitus patients. Turk J Biochem 2004; 29: 237-242.
- 12. Khan A, Safdar M, Khan MMA, Khattak KN, Anderson RA. Cinnamon Improves Glucose and Lipids of People With Type 2 Diabetes. Diabetes Care. 2003; 26:3215–3218.
- 13. Bruneton J. Pharmacognosy, Phytochemistry, Medicinal Plants. Lavoisier printing press, Paris. 1995; 509.
- Karnick CR. Pharmacopoeial Standards of Herbal Plants. Sri Satguru printing press, Delhi. 1994; 1138p.
- Kim SH, Hyun SH, Choung SY. Anti-diabetic effect of cinnamon extract on blood glucose in db/db mice. Journal of Ethnopharmacology. 2006; 104: 119-123
- Kitazuru ER, Moreira AVB, Mancini-Filho J, Delince'e H, Villavicencio ALCH. Effects of irradiation on natural antioxidants of cinnamon (*Cinnamomum zeylanicum N.*). Radiation Physics and Chemistry. 2004; 71; 37–39.
- 17. Shan B, Cai YZ, Sun M, Corke H. Antioxidant capacity of 26 spices extracts and characterization of their phenolic constituents. J. Agric Food Chem. 2005; 53: 7749-59.
- 18. Blomhoff R. Antioxidants and oxidative stres. Tidsskr Nor Laegeforen. 2004; 124: 1643-5.
- 19. Lopez P, Sanchez C, Batlle R, Nerin C. Solid- and vapour-phase antimicrobial activities of six essential oils: Susceptibility of selected food bourne bacterial and fungal strains. J. Agri Food Chem. 2005; 53: 6939-46.

- 20. Wichtl M, Bisset, NG. Herbal Drugs and Phytopharmaceuticals. Medpharm Scientific printing press. Stuttgart. 1994; 386p.
- Moretti N, Rabini RA, Nanetti L, Grechi G, Curzi MC, Cester N, Tranquilli LA, Mazzanti L. Sialic acid content in erythrocyte membranes from pregnant women affected by gestational diabetes. Metabolism. 2002; 51: 605-608.
- 22. Mazzanti L, Rabini RA, Testa I, Bertoli E. Modifications induceed by diabetes on the physicochemical and functional properties of erythrocyte plasma membrane. European J. Clin. Invest. 1989; 19: 84-89.
- 23. Flecha FG, Bermudez MC, Cedola NN, Gagliardino JJ, Rossi JP. Febs Lett 1990; 244: 484-486.
- 24. Ames BN. Asay of inorganic phosphate. Methods in Enzymology. Academic Press. 1966; 8: 115.
- 25. Yagi K. Assay for blood plasma or serum. Methods in Enzymolgy. 1984; 105: 328-337.
- Jain SK. Hyperglycemia Can Cause Membrane Lipid Peroxidation and Osmatic Fragility in Human Red Blood Cells. Journal of Biological Chemistry. 1989; 264: 21340-21345.
- 27. Lowry O, Rosenbraugh N, Farr L, Randall R. Protein measurement with theophilin-phenol reagent. J. Biol. Chem. 1951;183: 265–275.
- 28. Verspohl EJ, Bauer K, Neddermann E. Antidiabetic effect of *Cinnamomum cassia* and *Cinnamomum zeylanicum in vivo* and *in vitro*. Phytother Res. 2005; 19: 203-6.
- 29. Nandhini TA, Anuradha CV. Inhibition of lipid peroxidation, protein glycosylation and elevtion of membrane ion pump activity by taurine in RBC exposed to high glucose. Clin Chim Acta 2003; 336: 129-135.